



Research Article

## IMPACT OF MEDICINAL PLANT EXTRACTS AGAINST BACTERIAL STRAIN IN THANJAVUR AREA, TAMIL NADU, INDIA

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### ABSTRACT

The present study was aimed to evaluate the growth inhibitory effect of *Hibiscus rosasinensis*, *Azadirachta indica*, *Ficus religiosa* and *Ocimum sanctum* leaves extracts on *Esherichia coli*, *Salmonella typhii*, *Staphylococcus aureus* and *Enterobacter aerogen*. Aquous plant extracts were tested against 4 bacteria. Gel diffusion method, were used in this investigation. The antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Esherichia coli* 15 mm (Mean value in Dia.), when compared with other medicinal plant extracts. *Ficus religiosa* plant extract was zone of inhibition 14 mm (Mean value in Dia.) against *Esherichia coli* and 12 mm (Mean value in Dia.) against *Salmonella typhii*, *Staphylococcus aureus* and *Enterobacter aerogen*. The *Enterobacter aerogen* was highly inhibition 9 mm (Mean value in Dia.) against *Ocimum sanctum* plant extract. At the same time *Hibiscus rosasinensis* plant extract was highly sensitive 14 mm (Mean value in Dia.) against *Staphylococcus aureus*. The present observation, the *Azadirachta indica* and *Ficus religiosa* medicinal plant extracts are having best control of antibacterial activity.

**Keywords:** Growth inhibitory, Plant extracts, Bacterial strain, Thanjavur district.

### INTRODUCTION

Plants have been used for the treatment of diseases all over the world before the advent of modern clinical drugs. Natural photochemical are known to contain substances that can be used for therapeutic purposes or as precursor for the synthesis of novel drugs. Nearly 50% modern drugs are of natural products origin and as such these natural products play an important role in drug development in pharmaceutical industry. Plants remain the most common source of antimicrobial agents (Bibitha *et al.*, 2002; Maghrani *et al.*, 2005). Many aromatic plants have been used traditionally in folk medicine as well as to extend the shelf life of foods, showing inhibition against bacteria, fungi and yeast (Hulin *et al.*, 1998). Biologically active compounds from natural sources have always been a great interest for scientists working in infectious diseases (Perumal Samy and Ignacimuthu, 2000). There is an essential need to discover new antimicrobial compounds with diverse chemical structures and novel mechanisms of action. Therefore, search for medicinal plants with potential secondary metabolites have been extensively investigated as a source of medicinal agents.

Drug resistance is a serious global problem, and spread of resistance poses additional challenges for clinicians and the pharmaceutical industry. Use of herbal medicines in the developed world continue to rise because they are rich source of novel drugs and their bioactive principles form the basis in medicine, nutraceuticals, pharmaceutical intermediates and lead compounds in synthetic drugs (De *et al.*, 2002; Ncube *et al.*, 2008). Screening medicinal plants for biologically active compounds offers clues to develop newer antimicrobial agents. These compounds after possible chemical manipulation provide new and improved drugs to treat the infectious diseases (Natarajan *et al.*, 2003; Shah *et al.*, 2006). Plant based products extracts are cheaper alternatives to the development of synthetic drugs.

The plant-derived medicines are based upon the premise that they contain natural substances that can promote health and alleviate illness. So returns to natural substances are an absolute need of our time (Swayamjot *et al.*, 2005; Kumar *et al.*, 2007). In the last few years a number of studies have been conducted to verify the

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effectiveness of plant extracts against bacterial infections (Prashanth *et al.*, 2006; Ung *et al.*, 2010).

*Azadirachta Indica* belongs to the family Meliaceae, commonly known as neem. It is used in traditional medicine as a source of many therapeutic agents. *A. indica* (leaf, bark and seed) are known to contain antibacterial, antifungal activities against different pathogenic microorganisms and antiviral activity against vaccinia, chikungunya, measles and coxsackie B viruses (Biswas *et al.*, 2002). Different parts of neem (leaf, bark and seed oil) have been shown to exhibit wide pharmacological activities including; antioxidant, antimalarial, antimutagenic, anticarcinogenic, antiinflammatory, antihyperglycaemic, antiulcer and antidiabetic properties (Talwar *et al.*, 1997). The biological activities are attributed to the presence of many bioactive compounds in different parts. The present study evaluated the individual and in combination growth inhibitory effect of 4 medicinal plant extracts against 4 bacteria.

## MATERIALS AND METHODS

Plants were collected between the month of June and July 2015 in the Thanjavur area Tamil Nadu, India. Plant leaves were initially dried in an airconditioned, dehumidified room, then further dried in an oven at ca. 40°C for a total of seven days, and then finally ground to a fine powder.

Antimicrobial activity test was determined by the Kirby-Bauer disc diffusion method (Bauer *et al.*, 1966). The Antimicrobial activity was tested against isolated 4 bacterial strains. The medicinal plants of *Hibiscus rosasinensis*, *Azadirachta indica*, *Ficus religiosa* and *Ocimum sanctum* leaves extract were tested by the disc diffusion method. The extracts were prepared by reconstituting with aqueous. The test microorganisms were seeded into respective Mueller - Hinton agar medium by spread plate method 10 µl (10 cells/ml) with the 24h cultures of bacteria growth in Mueller - Hinton agar broth. After solidification the filter paper discs (5 mm in diameter) impregnated with the extracts were placed on test organism-seeded plates., *Escherichia coli*, *Salmonella typhi*, *Staphylococcus aureus* and *Enterobacter aerogen* were used for antibacterial test. Erythromycin (10 µg mlG1) used as positive control. The antibacterial assay plates were incubated at 37°C for 24h. After incubation, the results were observed and measured the diameter of inhibition zone (mm) around the each well.

## RESULTS AND DISCUSSION

The antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Escherichia coli* 15 mm (Mean value in Dia.), when compared with other medicinal plant extracts. *Ficus religiosa* plant extract was zone of inhibition 14 mm (Mean value in Dia.) against *Escherichia coli* and 12 mm (Mean value in Dia.) against *Salmonella typhi*, *Staphylococcus aureus* and *Enterobacter aerogen*. The *Enterobacter aerogen* was highly inhibition 9 mm (Mean value in Dia.)

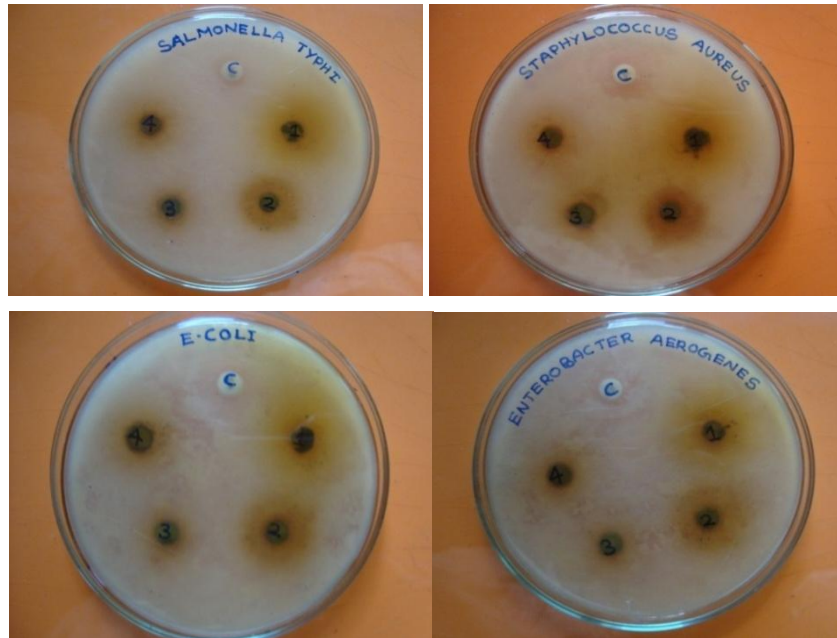
against *Ocimum sanctum* plant extract.

At the same time *Hibiscus rosasinensis* plant extract was highly sensitive 14 mm (Mean value in Dia.) against *Staphylococcus aureus*. The *Azadirachta indica* and *Ficus religiosa* medicinal plant extracts are having best control of antibacterial activity.

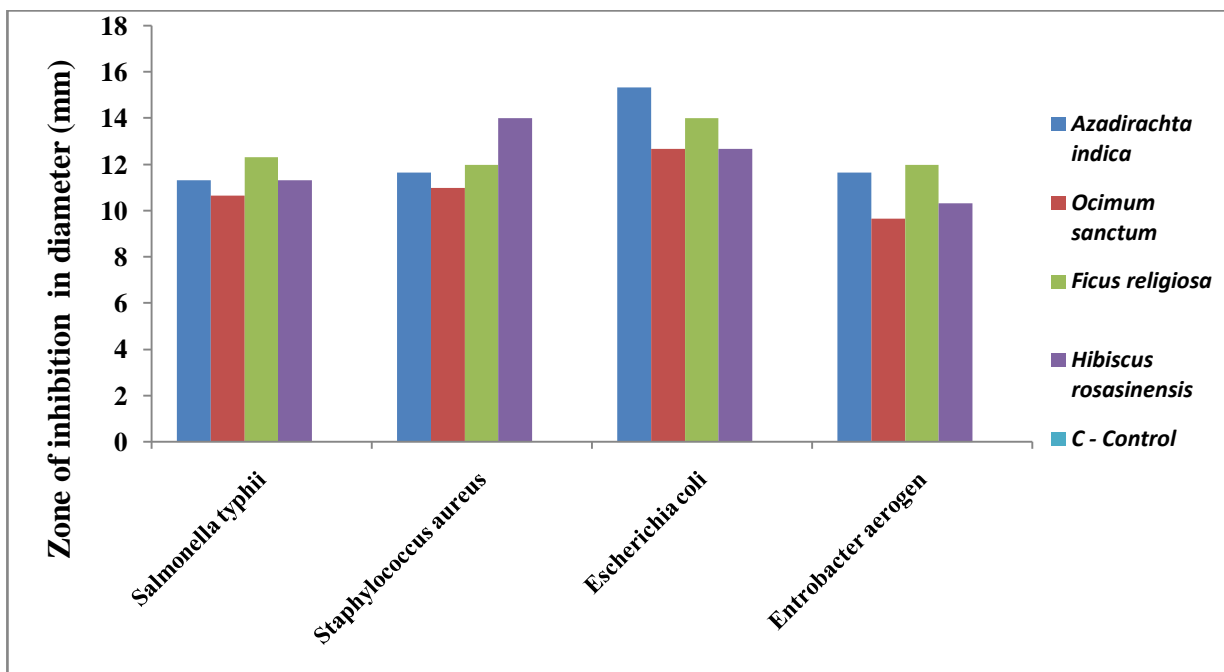
Antimicrobial activity test against five different medicinal plants (leaves) extracts such as *Hibiscus rosasinensis*, *Azadirachta indica*, *Ficus religiosa* and *Ocimum sanctum* (Leaves) were tested against some pathogenic bacteria such as *Escherichia coli*, *Salmonella typhi*, *Staphylococcus aureus* and *Enterobacter aerogen*, which were isolated from *Cirrhinus mrigala*. The antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Escherichia coli*, when compared with other medicinal plant extracts. According to Abalaka *et al.*, (2012) were studied the antibacterial effects of *A. indica* on the test organisms revealed that *P. aeruginosa* showed the highest zones of inhibition (mm) followed by *S. aureus* while *E. coli* had the least zone of inhibition (mm) at various extract concentrations of 500 mg/ml, 50 mg/ml and 5mg/ml. The extracts of *A. indica* showed a higher value of zones of inhibition on the tested organisms (Plate 1 and Figure 1). In a similar study hexane and aqueous extract of *Azadirachta indica*, inhibited *Escherichia coli*, *P. aeruginosa*, *S. pyogenes* and *S.aureus* (El-Mahmood *et al.*, 2010).

The methanol extract of the leaves of *Azadirachta indica* exhibited pronounced activity (28mm) against *Bacillus subtilis*, high activity (18mm) against the Gram-positive *Staphylococcus aureus* and the Gram-negative organisms *Proteus vulgaris* (18 mm) and *Salmonella typhi* (20 mm), low activity (14 mm) against *Pseudomonas aeruginosa* and inactive against *Escherichia coli* were reported by Nishant Rai *et al.*, (2011). The methanol extract of *Hibiscus* has got phytomedicine property it may be due to the nature of biologically active compounds present in *hibiscus* whose activity are enhanced in the presence on methanol and also methanol has a stronger extraction capacity which could have produced greater number of active constituents responsible for antibacterial activity (Barker *et al.*, 1995).

Plant based antimicrobial compounds have enormous therapeutic potential as they can serve the purpose without any side effects that are often associated with synthetic antimicrobials. The methanol, ethanol, ethyl acetate and chloroform and aqueous extracts of the leaves of *A. aspera*, *A. parviflora*, *A. indica*, and *C. odorata* were subjected to a preliminary screening for antimicrobial activity against two human pathogenic bacteria *E. coli* and *S. aureus*. High activity against the Gram-positive organism *E. coli* was found in aqueous and all tested solvent extracts of *A. indica*. In case of human pathogenic *S. aureus*, maximum inhibition of 8 mm was obtained in aqueous extracts of *A. indica*. Similar observations were reported from nimbolide isolated from neem seed oil showing antibacterial activity against *S. aureus* and *Staphylococcus coagulase* (Nazma and Rao, 1977).



**Plate 5.** Antimicrobial activity test against isolated bacteria from infected Fresh water carp *Cirrhinus mrigala*.



**Figure 14.** Antimicrobial activity test against isolated bacteria from infected carp *Cirrhinus mrigala*.

**Table 1.** Statistical analysis of mean and standard deviation in antimicrobial activity test in medicinal plants extract against bacteria.

Name of the Species	Zone of inhibition ( dia in mm)			
	S1. <i>Hibiscus rosasinensis</i>	S2. <i>Azadirachta indica</i>	S3. <i>Ficus religiosa</i>	S4. <i>Ocimum sanctum</i>
<i>Salmonella typhi</i>	11.33 ± 0.58	11.33 ± 1.15	12.33 ± 2.33	10.67 ± 2.08
<i>Staphylococcus aureus</i>	14.00 ± 3.00	11.67 ± 1.53	12.00 ± 3.60	11.00 ± 3.60
<i>E-coli</i>	12.67 ± 2.52	15.33 ± 2.52	14.00 ± 2.00	12.67 ± 2.52
<i>Enterobacter aerogen</i>	10.33 ± 0.58	11.67 ± 0.58	12.00 ± 4.58	9.67 ± 3.51

**CONCLUSION**

In the present investigation, the antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Esherichia coli* 15 mm (Mean value in Dia.), when compared with other medicinal plant extracts. *Ficus religiosa* plant extract was zone of inhibition 14 mm (Mean value in Dia.) against *Esherichia coli* and 12 mm (Mean value in Dia.) against *Salmonella typhii*, *Staphylococcus aureus* and *Enterobacter aerogen*. The *Azadirachta indica* and *Ficus religiosa* medicinal plant extracts were exhibited higher antibacterial activity.

**REFERENCES**

- Abalaka, M., Oyewole, O.A. and Kolawole, A.R., 2012. Antibacterial activities of *Azadirachta indica* against some bacterial pathogens, *Adv. Life Sci.*, 2(2), 5-8.
- Barker, J. T., Borris, R.P. and Carte, B., 1995. Natural product drug discovery and development: New perspective on international collaboration *J. Nat. Prod.*, 58, 1325-1357.
- Bauer, A.W., Kirby, W.M.M., Sherrsis, J.C., and Turk, M., 1966. Antibiotic susceptibility testing by a standardized single disc method. *Am. J. Clin. Pathol.*, 45(4), 163-182.
- Bibitha, B., Jisha, V.K., Salitha, C.V., Mohan, S. and Valsa, A.K., 2002. Antibacterial activity of different plant extracts. Short communication. *Indian J. Microbiol.*, 42, 361-363.
- Biswas, K., Ishita, C., Ranajit, K.B. and Uday, B., 2002. Biological activities and medicinal properties of Neem (*Azadirachta indica*). *Curr. Sci.*, 82, 1336-1345.
- De, N.B. and Ifeoma, E., 2002. Antimicrobial effects of components of the bark extracts of neem (*Azadirachta indica* A. Juss). *J. Technol. Dev.*, 8, 23-28.
- El-Mahmood, A.M., Ogbonna, O.B. and Raji, M., 2010. The antibacterial activity of *Azadirachta indica* (neem) seeds extracts against bacterial pathogens associated with eye and ear infections, *J. Med. Plants Res.*, 4(14), 1414-1421.
- Hulin, V., Mathot, A.G., Mafart, P. and Dufosse, L., 1998. Les proprietes anti-microbiennes des huiles essentielles et composees d'aromes. *Sci Aliments*, 18: 563-582.
- Kumar, K., Devis, S.S., Krishnamurthi, K., Kanade, G.S. and Chakrabarti, T., 2007. Enrichment and isolation of endosulfan degrading and detoxifying bacteria. *Chemosphere*, 68, 317-22.
- Maghrani, M., Zeggwah, N., Michel, J. and Eddouks, M., 2005. Antihypertensive effect of *Lepidium sativum* in spontaneously hypertensive rats. *J. Ethnopharm.*, 102(1-2), 193-197.
- Natarajan, V., Veugopal, P.V. and Menon, T., 2003. Effect of *Azadirachta indica* (neem) on the growth pattern of dermatophytes. *Indian J. Med. Microbiol.*, 21, 98-101.
- Nazma, B.S.V. and Rao, J.M., 1977. Antifungal Activity of Gedunin. *Curr. Sci.*, 46, 714-715.
- Ncube, N.S., Afolayan, A.J. and Okoh, A., 2008. Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *African J. Biotechnol.*, 7, 1797-1806.
- Nishant Rai, Aditi Grover and Bhandari, B.S., 2011. Antimicrobial Activity of Medicinal plants- *Azadirachta indica*, *Allium cepa* L. and *Aloe vera* L. *Int. J. Pharm. Tech. Res.*, 3(2), 1059-1065.
- Perumal Samy, R. and Ignacimuthu, S., 2000. Antibacterial activity of some medicinal plants from Eastern Ghats, South India, Solai bull. *J. Ethnopharmacol.*, 72, 39-41.
- Prashanth, K., Neelam, S., Harish, P. and Rajani, M., 2006. Search for antibacterial and antifungal agents from selected Indian medicinal plants. *J. Ethnopharmacol.*, 107, 182-188.
- Shah, J.S., Shah, M.B., Goswami, S.S. and Santani, D.D., 2006. Mechanism of action of antiulcer activity of bark extracts of *Manikara hexandra* against experimentally induced gastric ulcers in rats. *Phcog. Mag.*, 2, 40-45.
- Swayamjot, K., Husheem, M., Saroj, A., Pirkko, L.H. and Subodh Kumar, K., 2005. The in vitro cytotoxic and apoptotic activity of Triphalaan Indian herbal drug. *J. Ethnopharmacol.*, 97, 15.
- Talwar, G.P., Raghuvanshi, P., Misra, R., Mukherjee, S. and Shah, S., 1997. Plant immunomodulators for termination of unwanted pregnancy and for contraception and reproductive health. *Immunol. Cell Biol.*, 75, 190-192.
- Choi, U.K., Lee, O.H., Lim, S.I. and Kim, Y.C., 2010. Optimization of antibacterial activity of *Perilla frutescens* var. *acuta* leaf against *Pseudomonas aeruginosa* using the evolutionary operation factorial design technique. *Int. J. Mol. Sci.*, 11(10), 3922- 3932.